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# **Regulatory News 2023**

New EN ISO 15213 series for the enumeration and detection of *Clostridium* spp. in the food chain

The International Organization for Standardization (ISO) is publishing the three-part EN ISO 15213 series, which specifies the enumeration and detection of sulfite-reducing *Clostridium* spp., including *C. perfringens*, in a broad range of foods, pet food and animal feed, in samples from the primary production stage and in environmental samples in food and feed production and handling.

# New EN ISO 15213-1:2023 at a glance ...

- Describes the horizontal method for the enumeration of *Clostridium* spp. by colony-count technique.
- Scope of the method has been changed from "sulfite-reducing bacteria" to "sulfite-reducing *Clostridium* spp." and includes samples from the primary production stage.
- Typical colonies on Iron sulfite agar (ISA) are confirmed by anaerobic growth and no growth under aerobic conditions.
- Concentration of sulfite in the Iron sulfite agar (ISA) has been reduced from 1.0 g/L to 0.5 g/L.
- Ten minute heat treatment at 80 °C for selection of spores has been made optional.
- Option to use tubes for inoculation and option to incubate the samples at 50 °C for enumeration of thermophilic sulfite-reducing bacteria have been removed.
- A special protocol for the enumeration of sulfitereducing *Clostridium* spp. in feed has been added in an informative annex.
- Performance characteristics of the method, determined in an interlaboratory study, have been added.
- The main technical changes are significant and have major impact on the performance characteristics of the method.

- Part 1: Enumeration of sulfite-reducing Clostridium spp. by colony-count technique.
   It has replaced ISO 15213:2003.
- Part 2: Enumeration of Clostridium perfringens by colony-count technique.
   It is set to replace EN ISO 7937:2004.
- Part 3: Detection of *Clostridium perfringens*. This part will be newly published.

# Part 1: Enumeration of *Clostridium* spp. acc. to new EN ISO 15213-1:2023

#### **Procedure step**

Sample preparation

- Preparation of test portion or initial suspension.
- OPTIONAL for selection of spores: Heat treatment at  $(80 \pm 2)$  °C for  $(10 \pm 1)$  min

Plating

- Prepare decimal dilutions.
- Transfer 1 mL of 1:10 diluted sample or decimal dilution into empty 90 mm Petri dish.

Plating (pour plate)

- Add 12-15 mL Iron sulfite agar (ISA) at 44-47 °C.
- After solidification, overlay with 5 mL of liquid ISA.
- Incubate under anaerobic conditions at (37  $\pm$  1) °C for (48  $\pm$ 2) h.

Enumeration and confirmation

- Count typical colonies of presumptive sulfitereducing Clostridium spp.:
   black or grey to yellow brownish colonies.
- For confirmation, take 5 colonies and streak out each colony onto two non-selective agar plates (e.g. Columbia blood agar).

From each pair of plates:
 Incubate 1 plate at 37 °C for 20 h ± 2 h aerobically.
 Incubate 1 plate at 37 °C for 20 h ± 2 h anaerobically.

Only colonies that grow ANAEROBICALLY but not aerobically belong to the genus  ${\it Clostridium}.$ 

This and other colonies with the same morphology on ISA are counted as sulfite-reducing *Clostridium* spp.

### Part 2: Enumeration of Clostridium perfringens acc. to new EN ISO 15213-2:2023

# New EN ISO 15213-2:2023 at a glance ...

- Describes the horizontal method for the enumeration of *Clostridium perfringens* by colony count technique.
- Scope has been expanded to include samples from the primary production stage.
- Typical colonies on Tryptose sulfite (TSC) agar are confirmed by SIM agar or Acid phosphatase test.
- Ten minute heat treatment at 80 °C has been made optional.
- The selective medium has been re-named from Sulfite-cycloserine agar (SC) to Tryptose sulfite cycloserine agar (TSC) without changes in the formulation.
- Description of Acid phosphatase test has has been aligned with ISO 14189.
- Molecular differentiation between pathogenic and non-pathogenic
   C. perfringens has been added in an informative annex.
- Performance characteristics of the method, determined in an interlaboratory study, have been added.
- The main technical changes are significant and have a major impact on the performance characteristics of the method.

#### Procedure step

Sample preparation

- Preparation of test portion or initial suspension.
- OPTIONAL for selection of spores: Heat treatment at  $(80 \pm 2)$  °C for  $(10 \pm 1)$  min

- Prepare decimal dilutions.
- Transfer 1 mL of 1:10 diluted sample or decimal dilution into empty 90 mm Petri dish.

**Plating** 

- Add 12-15 mL Tryptose sulfite cycloserine (TSC) agar at 44-47 °C.
- After solidification, overlay with 5 mL of liquid TSC agar.
- Incubate under anaerobic conditions at (37  $\pm$  1) °C for (20  $\pm$  2) h.



- Count typical colonies of presumptive Clostridium perfringens:
- black or grey to yellow-brownish colonies.
- For confirmation, take 5 colonies and subculture these on a non-selective agar (e.g. Columbia blood agar).
- Incubate under anaerobic conditions at (37  $\pm$  1) °C for (20  $\pm$  2) h.
- Confirm colonies are *C. perfringens* by using either Acid phosphatase or SIM agar test.

Enumeration & confirmation

- For Acid phosphatase test:
- Spread colonies on filter paper and add 2-3 drops of acid phosphatase reagent.

A purplish color developing within 3-4 min is considered a positive reaction.

- For SIM agar test:
- Stab colonies from non-selective agar plates into SIM tubes.

Incubate under anaerobic conditions at (37  $\pm$  1) °C for (22  $\pm$  2) h with loose caps.

Tubes showing blackening (sulfite production: positive), NO growth outside the inoculation stab (motility: negative) and NO red ring formation after adding Kovac's reagent (indole production: negative) are confirmed as positive for *C. perfringens*.

## Part 3: Detection of Clostridium perfringens acc. to new EN ISO/TS 15213-3:2023

Step

3

**Procedure step** 

# New EN ISO/TS 15213-3:2023 at a glance ...

- Describes the horizontal method for the detection of *Clostridium perfringens*.
- Scope includes the detection of C. perfringens in food, feed, and environmental samples in food and feed production and handling, and for samples from the primary production stage.
- Rapid Perfringens Medium (RPM) is described as a new culture medium for selective enrichment.
- For isolation, Tryptose sulfite (TSC) agar and new Lactose egg-yolk neomycin agar (LENA) are used as described.
- Typical colonies on TSC agar and/or LENA are confirmed by SIM agar or Acid phosphatase test.
- Molecular differentiation between pathogenic and non-pathogenic
   C. perfringens has been added in an informative annex.
- Performance characteristics of the method, determined in an interlaboratory study with some participating laboratories, have been added.

### Liquid sample or initial suspension + Rapid perfringens medium (RPM): Sample preparation Tenfold dilution (of 1 mL or 10 mL sample / initial suspension) (with RPM). Selective - Incubate at (46 $\pm$ 1) °C for (18 $\pm$ 4) h. enrichment - Streak 10 $\mu L$ onto the surface of TSC agar and 10 μL onto the surface of LENA plates. - Incubate TSC agar plates for (24 ± 2) h Plating at $(37 \pm 1)$ °C anaerobically. - Incubate LENA plates for (24 $\pm$ 2) h at (46 $\pm$ 1) °C anaerobically. - Typical colonies of presumptive *C. perfringens* on TSC agar: black or grey to yellow-brownish colonies: on LENA: yellow colonies (acid fermentation of lactose) with precipitation (lecithinase reaction). - For confirmation, take 5 colonies and subculture these on a non-selective agar (e.g. Columbia blood agar). - Incubate under anaerobic conditions at (37 ± 1) °C for $(20 \pm 2) h$ . - Confirm colonies are *C. perfringens* by using either acid phosphatase or SIM agar test. - For acid phosphatase test: Confirmation Spread colonies on filter paper and add 2-3 drops of Acid phosphatase reagent. A purplish color developing within 3-4 min is considered a positive reaction. - For SIM agar test: Stab colonies from non-selective agar plates into SIM Incubate under anaerobic conditions at $(37 \pm 1)$ °C for $(22 \pm 2)$ h with loose caps. Tubes showing blackening (sulfite production: positive), NO growth outside the inoculation stab (motility: negative) and NO red ring formation after adding Kovac's reagent (indole production: negative) are confirmed as positive for C. perfringens.

As a worldwide leading provider of a broad range of granulated and readyto-use culture media for food, beverage and water microbiology, we closely track and support the development of relevant standards aimed at increasing consumer confidence and safety.

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### Compliance with the new EN ISO 15213 series

We are implementing all the requirements as described in the new EN ISO 15213 series.

For more information, please visit our webpage SigmaAldrich.com/updates-on-iso-standards

## Dehydrated culture media - from basic to prime

To best suit your needs, we offer two media formats: superior granulated GranuCult® and powdered NutriSelect® culture media. The added designations basic, plus or prime indicate the quality control level, from basic quality control, to standard QC plus, to prime for full regulatory compliance.

### **Ordering Information**

Product description	Pack size	Cat. No.
GranuCult® prime Iron Sulfite Agar (ISA) acc. ISO 15213-1	500 g	1108640500
GranuCult® prime Columbia agar (base) acc. ISO 10272 and EP/USP/JP	500 g	1002140500
GranuCult® prime TSC (Tryptose Sulfite Cycloserine) Agar (base) acc. ISO 15213, ISO 14189 and FDA-BAM	500 g	1119720500
TSC (Cycloserine) Selective supplement	1 x 10 vials	1008880010
GranuCult® prime Rapid Perfringens Medium (RPM) acc. ISO 15213-3	500 g	1167510500
GranuCult® prime SIM (Sulfite Indole Motility) Agar acc. ISO 15213	500 g	1054700500
Bactident® Indole (Kovac's indole reagent) acc. ISO and FDA-BAM	30 mL	1113500001
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Anaerotest® strips for microbiology (to indicate an anaerobic atmosphere)	50 strips	1323710001
Anaerocult® A mini (gas generator system for 1-4 Petri dishes)	25 x 1 set	1323690001
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